This article was downloaded by: On: 24 January 2011 Access details: Access Details: Free Access Publisher Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Macromolecular Science, Part A

Publication details, including instructions for authors and subscription information: http://www.informaworld.com/smpp/title~content=t713597274

Benzo[h]quinoline based Macrocyclic Copper(II), Cobalt(II) Complexes: Synthesis, Characterization and Light induced DNA Cleavage Studies

H. R. Prakash Naik^a; H. S. Bhojya Naik^a; D. S. Lamani^a; T. Aravinda^a; B. Vijaya Kumar^a; B. Vinay Kumar^a; M. Yogesh^a; N. Sharath^a; P. N. Prashanth Kumar^a

^a Department of Studies and Research in Industrial Chemistry, School of Chemical Sciences, Kuvempu University, Shankaraghatta, India

To cite this Article Naik, H. R. Prakash , Naik, H. S. Bhojya , Lamani, D. S. , Aravinda, T. , Kumar, B. Vijaya , Kumar, B. Vinay , Yogesh, M. , Sharath, N. and Kumar, P. N. Prashanth(2009) 'Benzo[h]quinoline based Macrocyclic Copper(II), Cobalt(II) Complexes: Synthesis, Characterization and Light induced DNA Cleavage Studies', Journal of Macromolecular Science, Part A, 46: 8, 790 – 795

To link to this Article: DOI: 10.1080/10601320903004558 URL: http://dx.doi.org/10.1080/10601320903004558

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: http://www.informaworld.com/terms-and-conditions-of-access.pdf

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

Benzo[h]quinoline based Macrocyclic Copper(II), Cobalt(II) Complexes: Synthesis, Characterization and Light induced DNA Cleavage Studies

H. R. PRAKASH NAIK, H. S. BHOJYA NAIK^{*}, D. S. LAMANI, T. ARAVINDA, B. VIJAYA KUMAR, B. VINAY KUMAR, M. YOGESH, N. SHARATH and P. N. PRASHANTH KUMAR

Department of Studies and Research in Industrial Chemistry, School of Chemical Sciences, Kuvempu University, Shankaraghatta-577 451, India

Received January 2009, Accepted March 2009

A new ligand dibenzo[h]quinolineno[1,3,7,9] tetraazacyclododecine-7,15 (14H, 16H)-dibenzene (L) and its Co(II)/Cu(II) metal complexes of type [MLX₂] (Where (M = Co(II) (5), Cu(II) (6) and X = Cl) were synthesized and are well characterized by FT-IR, ¹H-NMR, FAB mass elemental analysis, and electronic spectral data. The role of the cobalt/copper metals in photo-induced DNA cleavage reactions was explored by designing complex molecules having macrocyclic structure. Finally, we have shown that photocleavage of plasmid DNA is more efficiently enhanced when this macrocyclic ligand is irradiated in the presence of copper(II) than that of cobalt metal.

Keywords: Benzo[h]quinoline-macrocycles, Co(II)/Cu(II) complexes, photo-induced DNA cleavage

1 Introduction

In recent years, the discovery of cisplatin for cancer treatment, numerous transition metal complexes have been synthesized and screened for their anticancer properties. One strategy in such efforts is to synergize the beneficial effect of the ligand and the activity of the metal to produce a complex with enhanced activity (1).

Development of macromolecular architectures possessing defined structural motifs is a focus of intense research in the area of molecular or ion recognition (2), supramolecular chemistry (3), and drug design (4). A fundamental step to realize structure-based interactions should be an appropriate design and synthesis of structural modules, which can be assembled to build tailored molecular structures. Currently, there is considerable interest in complexes of polydentate macrocyclic ligands because of the variety of geometrical forms available and the possible encapsulation of the metal ion (5).

Deoxyribonucleic acid (DNA) is the primary target molecule for most anticancer and antiviral therapies according to cell biology. Investigations of the interaction of DNA with small molecules are basic work in the design of new types of pharmaceutical molecules. Since the chemical nuclease activity of the copper and cobalt complexes were discovered in 1980s (6-8), studying the interaction model and the mechanism of transition metal complexes with DNA, and exploring the application of metal complexes in antineoplastic medication, molecular biology and bioengineering were hotspots in recent year. When some kinds of metal complexes interacted with DNA, they could induce the breakage of DNA strands by appropriate methods. Thus, after the double DNA strands are broken, the replication ability of cancer gene is destroyed. The interactions of metal complexes with DNA constitute a significant area of research which has attracted considerable attention from both inorganic chemists and biochemists because studies have shown that they are related to the development of new DNA reagents for biotechnology and medicine (9, 10). In recent years, binding studies of transition metal complexes have become very important in the development of DNA molecules probes and chemotherapeutics (11 - 13).

In view of above vast chemotherapeutic applications and in continuation of quest on quinoline based macrocycles (14). Here, we describe for the first time cobalt/coppermacrocyclic complexes of benzo[h]quinoline moiety and their DNA nuclease activity.

^{*}Address correspondence to: H. S. Bhojya Naik, Department of Studies and Research in Industrial Chemistry, School of Chemical Sciences, Kuvempu University, Shankaraghatta-577 451, India. Fax: +91-8282-256255; E-mail: hsb_naik@rediffmail.com

2 Experimental

2.1 Chemicals

All chemicals used for the synthesis were of analytical grade and procured from Sigma Chemical Co., U.S.A., E. Merck, Germany, Sarabhai Merck Company, India and ophenylenediamene was purchased from S. D. Fine Chemicals Pvt. Ltd. The TLC was performed on Baker-Flex silica gel 1B-F (1.55) plates in the following solvent systems: ethyl acetate and petroleum ether (8:3). Melting points were determined on a Mel-Temp apparatus and are uncorrected. IR spectra were recorded in the matrix of KBr with a Perkin-Elmer 1430 spectrometer. ¹H-NMR spectra were recorded on a Jeol spectrometer (400 MHz), and chemical shifts (δ) are given in ppm relative to the signal for TMS as internal standard. C, H and N analyses were performed at Cochin University, Sophisticated Test and Instrumentation Center, Kochi, Kerala, India. Conductivity measurements were determined in DMF (10^{-3} M) using an ELICO-CM82 Conductivity Bridge.

2.2 Preparation of N,N'-bis[(Z)-(2-chlorobenzo[h]quinolin-3-yl)methylidene]ethane-1,2-dibenzene (3)

The ethanolic solution of 2-chloro-3-formyl-benzo[h] quinoline (9.64 g, 0.04 mol) and o-phenylenediamine (2.16 g, 0.02 mol) (25 ml each) in 2:1 molar ratio was refluxed for 3–4 h. A yellowish product separates out and was washed with cold ethanol, dried under vacuum, and recrystallized from an ethyl acetate/dichloromethane solvent system. Yield 94%, m.p. 156–158°C.

2.3 Preparation of Dibenzo[h]quinolineno[1,3,7,9] tetraazacyclododecine-7, 15 (14H, 16H)-dibenzene (L)

The compound **3** (5.55 g, 0.01 mol) was dissolved in DMF (30 ml) and added to 25 ml o-phenylenediamine (1.08 g, 0.01 mol) in 1:1 molar ratio. The solution was refluxed in the presence of anhydrous potassium carbonate (2.76 g, 0.02 mol) as catalyst for 10–12 h. The reaction was monitored by TLC using petroleum ether and ethyl acetate (8:3) as eluent. A greenish white precipitate was separated in ice cold water. The resulting product was collected by filtration, washed with cold water, dried under vacuum, and recrystallized from ethanol. Yield 81% m.p. 248–250°C, FT-IR cm⁻¹ 3462 (-NH-); 2938 (Ar-CH); 1626 (C=N); (other peaks) 1502, 1295, 1105, 958, 785, 675. ¹H-NMR(CDCl₃) δ : 8.96 (2s, 2H, NH); 8.87 (2d, 2H, CHN, D₂O exchangeable proton); 7.0–7.9 (m, 22H, Ar-H, Quinoline and ophenylenediamine).

2.4 General Procedure for the Preparation of Complexes

A simple method has been adopted for the preparation of the complexes. The hot ethanol solution of ligand (L) and hydrated metal salt in 1:1 molar ratio were mixed. The mixture was refluxed for about 3–4 h, at $80 \pm 5^{\circ}$ C, the obtained residue was recrystallized from ethanol. Various attempts to develop the crystals suitable for X-ray diffraction studies such as slow diffusion, crystallization using mixtures of solvents and low temperature crystallization were unsuccessful.

2.5 [Co(L)Cl₂]nH₂O: Cobalt(II) Complex with ligand(L) dibenzo[h]quinolineno[1,3,7,9] tetraazacyclododecine-7,15 (14H, 16H)-dibenzene

Ligand (L) was dissolved in (25 ml) ethanol and added to the hot ethanolic solution of cobalt (II) chloride (25 ml) in 1:1 molar ratio under boiling conditions and refluxed for 3–4 h (Scheme 1). A blue colored precipitate formed was collected by filtration and dried. Similarly, the same procedure was followed for Cu (II) complex, and the experimental data were summarized in Table 1.

2.6 DNA Photocleavege Cleavage Experiments

The experiments were performed in a volume of 2 ml containing pUC19 DNA in 5 μ mol/L phosphate buffer contained 10 μ mol/L NaCl, pH 7.4, in the presence of different concentrations (200–600 μ mol/L) of complexes. Immediately prior to irradiating the samples with UV light, H₂O₂ was added to a final concentration of 2.5 μ mol/L.

The reaction volumes were held in caps of polyethylene microcentrifuge tubes, which were placed directly on the surface of a trans-illuminator (8000 mW/cm) at 360 nm. The samples were irradiated for 5 min at room temperature. After irradiation, 0.5 ml of a mixture containing 0.25% bromophenol blue, 0.25% xylene cyanol FF, and 30% glycerol was added to the irradiated solution. The samples were then analyzed by electrophoresis on a 1% agarose horizontal slab gel in Tris-borate buffer (45 μ mol/L Tris-borate, 1 μ mol/L EDTA). Untreated pUC19 DNA was included as a control in each run of gel electrophoresis, which was carried out at 1.5 V/cm for 15 h. Gel was stained in ethidium bromide (1 mg/ml) and photographed under UV light (15).

3 Results and Discussion

3.1 Chemistry

A novel dibenzo[h]quinolineno[1, 3, 7, 9] tetraazacyclododecine-7,15 (14H, 16H)- dibenzene, macrocyclic ligand (L) has been synthesized in two steps as per Scheme 1. In the first step, the o-phenylenediamine (2) reacts with 2chloro-3-formylbenzo[h]quinoline (1), in 1:2 molar ratio in ethanol, a brownish yellow colored product N-[-(2-chlorobenzo[h]quinolin-3-yl) methylene]-N-(2-chlorobenzo[h] quinolin-3-yl) methylene] benzene-1,2-diamine separated



Sch. 1. Synthetic pathway for the macrocyclic dibenzo[h]quinolineno[1, 3, 7, 9] tetraazacyclododecine-7,15 (14*H*, 16H)-dibenzene ligand (L).

out. In a second step, it reacts with o-phenylenediamine in 1:1 molar ratio in DMF solvent, gave a greenish colored solid. The TLC has established the purity of the compound by dissolving the ligand in ethanol using petroleum ether and ethyl acetate (8:3) as eluent. One spot was observed in the TLC plate after developing in an iodine chamber indicating that the compounds were pure. The formation of this macrocyclic molecule was confirmed by the results of FT-IR and resonance peaks, ¹H-NMR and elemental analysis data's. Accordingly, these new macrocyclic complexes of the type [MLX₂], were synthesized by the reaction of the ligand (L) with the corresponding Co/Cu metal salts in 1:1 molar ratio in ethanol solution. The formation of the complex may be represented by the following reaction:

$$CoX_2.H_2O + L \rightarrow Co(L)X_2 + nHlO$$

 $CuX_2.H_2O + L \rightarrow Cu(L)X_2 + nH_2O$

The complexes are microcrystalline in nature and found to be soluble in most of the organic solvents. The elemental analysis data shows that the complexes have a composition of $[Co(L)Cl_2]$, $[Cu(L)Cl_2]$. The magnetic moment

 Table 1. Analytical and physical properties of the metal complexes dibenzo[h]quinolineno[1, 3, 7, 9] tetraazacyclododecine-7,15 (14H, 16H)-dibenzene (L)

Complex	Color	Molecular Wt (Yield %)	$m.p.^{\circ}C$	Meff (B.M)	$(\Delta m \ \Omega^{-1} \ cm^{-1} mol^{-1})$	Elemental analysis Calcd. (Found %)
$(4) C_{40}H_{26}N_6$	Greenish	590.67 (81)	248		_	C: 81.34 (80.95) H: 4.44 (4.32) N: 14 23 (14 01)
$\begin{array}{l} [Co(L)Cl_2](5) \\ C_{40}H_{28}N_6Cl_2CoN_6 \end{array}$	Dark blue	722.53 (78)	>255	2.56	72	C: 66.49 (66.39) H: 3.91 (3.84) N: 11.63 (11.55)
$\begin{array}{c} [Cu(L)Cl_2] \ (6) \\ C_{24}H_{22}N_6Cl_2Cu \end{array}$	Dark Red	727 (75)	>260	2.02	75	C: 66.07 (65.94) H: 3.88 (3.81) N: 18.35 (18.41) Cu: 8.74 (8.71)



Fig. 1. Three Dimensional (3D) structure of Co(II) (5)/Cu(II)(6) Macrocyclic complex.

value 2.56 for Co(II), and 2.02 for Cu(II), which are greater than spin-only value 1.75 (B.M) and hence, paramagnetic in nature, exhibits high–spin octahedral geometry. The coordination spheres of complexes, similar to those of Nickel (II)-type macrocyclic complexes, have been reported to be sixcoordinate octahedral geometry (16). Hence, in the present studies, the experimental results suggest that the title complexes possess octahedral geometry. Molar conductivity was studied in DMF, the range of 72–75 Δm Ω^{-1} cm⁻¹ mol⁻¹ indicating that both the complexes are 1:1 electrolytes and may be formulated as [MLX₂]. In addition proposed three dimensional (3D) structures of both copper and cobalt macrocyclic complexes were shown in Figure 1.

3.2 FT-IR Spectra

IR spectra of complexes were recorded in the matrix of KBr pellets with a Perkin-Elmer 1430 spectrometer. The absence of bands corresponding to the amino groups of etheylenediamene and carbonyl groups of aldehydic 2-chloro-3-formyl-quinoline, suggests the formation of the proposed macrocyclic ligand (L). Further, the two intensive bands at 1626 cm⁻¹ and 3462 cm⁻¹ assignable to uncoordinated ν (C=N) and ν (N-H) of amine group, respectively confirms the proposed structure (17,18). In addition, the formation of macrocyclic structure was conformed by its ¹H-NMR spectra. However, IR spectra of complexes derived from the ligand (L) shows a slight shift to the lower frequency in ν (C=N) and appeared in the region 1599–1621 cm⁻¹ suggesting its coordination with metal ion. In addition, a strong characteristic band of ν (-NH-) appeared at

3176 cm⁻¹, and bands at 1452–1421 cm⁻¹ for all the complexes correspond to C–H binding vibrations, respectively. The appearance of new medium–intensity bands in the region 762–769 cm⁻¹ in the macrocyclic complexes may be assigned to ν (M-N) vibrations. The bands at 451–478 cm⁻¹ were assigned to ν (M-Cl) vibrations, and the values are summarized in Table 2.

3.3 ¹H-NMR Spectra

The ¹H-NMR spectra were recorded on a Jeol spectrometer (400 MHz), and chemical shifts (δ) are given in ppm relative to the signal for TMS as the internal standard. The absence of proton resonance signals of free NH₂ and aldehydic (CHO) groups indicates the condensation between amine and carbonyl group of aldehydic 2-chloro-3-formylquinoline. The ¹H-NMR spectra of the ligand recorded in CDCl₃ show a doublet at δ : 8.87 ppm (d, 1H, CHN D₂O exchangeable), may be due to hydrogen bonding and anisotropy effect of the adjacent and other aromatic resonated protons, and signal exhibits singlate at δ : 8.96 ppm (2s, 2H, NH) was ascribed. The multiplet signals attributed at 7.0–7.9 ppm are due to (m, 22H, Ar-H) aromatic quinoline moiety.

3.4 The pUC 19 DNA Cleavage Studies

Supercoiled plasmid DNA cleavage by the Co(II)/Cu(II) complexes and their analogues was studied in the presence of H_2O_2 or any reducing agents (Fig. 2) and a time/concentration dependent cleavage was observed. We

Table 2. Characterizations of IR cm⁻¹ bands of ligands and their metal complexes

Compound	v(N–H)	v(Ar-CH)	v(C–N)	v(C–C)	v (С–Н)	v(M-N)	v(M-Cl)
$\frac{1}{(4) C_{40} H_{26} N_6}$	3462s	2938 m	1626s	1485 m	1454s	_	_
$[Co(L)Cl_2](5) C_{40}H_{28}N_6Cl_2CoN_6$	3182s	2851 m	1611s	1457 m	1417s	762s	478 m
$[Cu(L)Cl_2]$ (6) $C_{24}H_{22}N_6Cl_2CuN_6$	3179s	2854 m	1595s	1458 m	1416s	768s	451 m



Fig. 2. Effects of Co/Cu complexes (5/6) at various concentrations (200-600 μ mol/L) on the pUC 19 supercoiled DNA against OH generated by photolysis at 360 nm in presence of H₂O₂. Lane 1, Untreated DNA (control); lane 2, DNA + H₂O₂; lane 3, DNA + Complex 5 (200 μ mol/L); lane 4, DNA + Complex 5 (400 μ mol/L); lane 5, DNA + Complex 5 (600 μ mol/L); lane 6, DNA + Complex 5 (800 μ mol/L); lane 7, DNA + Complex 6 (200 μ mol/L); lane 8, DNA + Complex 6 (400 μ mol/L); lane 9, DNA + Complex 6 (600 μ mol/L); lane 10, DNA + Complex 6 (800 μ mol/L).

found that the supercoiled DNA (form I) was cleaved by **5** or **6** only after 1 and 2 h. The cleavage activity of **6** is considerably more than **5**. In order to clarify the DNA cleavage mechanism, complexes **5** and **6** were investigated in the presence of chelating agent. Both the complexes were tested for DNA cleavage under hydrolytic conditions, and a concentration dependent cleavage was observed. Reaction that leads to formation of open circular DNA (form II) from the supercoiled from I over various concentrations of complexes **5/6** (100–600 μ M/L) and constant DNA concentration was followed for different concentration at 37°C (Fig. 1).

It is now recognized that the extremely reactive OH radical derived from O_2^- and H_2O_2 is a cause of DNA strand scission in cellular damage (19). Figure 1 shows the electrophoretic pattern of DNA after UV-photolysis of H_2O_2 (2.5 μ mol/L) in the absence or presence of the complex.

The faster-moving band corresponds to the native form of supercoiled circular DNA (scDNA) and the slowermoving band being the open circular form (ocDNA). The UV irradiation of DNA in the presence of H_2O_2 (lane 2) caused the cleavage of scDNA to give open coiled DNA (ocDNA) and the linear form (linDNA), indicating that · OH generated by UV-photolysis of H_2O_2 produced DNA strand scission. The presence of the complexes under investigation increases the DNA damage which has been particularly implicated in carcinogenesis (20).

In the present case, complex **6** is more effective than complex **5** to undergo a single cleavage event. So the distinct cleavage efficiencies observed for the present complexes can be rationalized by their significantly different coordination geometries. Indeed, it has been shown that the coordination geometry plays an important role in Cu(II)/Cu(I) redox processes (21). Both complexes which are octahedral, may easily accommodate an electron in its co-planar d(x2-y2) orbital (22). Accordingly, which may likely to occur with a Fenton-type mechanism. It has been demonstrated that the cleavage of DNA in the absence of a reductant is possible with copper(II) complexes through an effective activation of molecular oxygen, generating reactive oxygen species (23). The favorable Cu(II) to Cu(I) redox potential is then coupled with a self-hydrogen abstraction from the DNA molecule (most probably from the sugar moieties). The occurrence of this process instigates a single DNA cleavage event, through a Fenton mechanism. This DNA cleavage may become catalytic if the ligands coordinated to the Cu ion facilitate the Cu(II)/Cu(I) cycle (24).

4 Conclusions

In summary, we describe the preparation and photoreactivity of a novel benzo[h]quinoline based macrocyclic Co(II)/Cu(II) transition metal complexes. The overall strategy of metal complex activation *via* LMCT excitation in the visible spectral region is operative. This paper reports that the Cu(II) macrocyclic complex can effectively cleavage DNA in mild condition *via* a non-oxidative mechanism than that of Co(II) macrocyclic complex. Based on our experiment results, we may predict that the Cu(II)-L complex would be another potential DNA hydrolytic cleavage agent.

Acknowledgements

One of the authors (H. R. Prakash Naik) thankful to Director NMR research institute, Indian Institute of Science (IISc) Bangalore for providing spectral facilities and Sophisticated Test & Instrumentation Center, Cochin University, Kochi, Kerala, India for elemental analysis. SC/ST cell Kuvempu University for providing Junior Research Fellowship (JRF).

References

- Christofis, P., Katsarou, M., Papakyriakou, A., Sanakis, Y., Katsaros, N. and Psomas, G. (2005) J. Inorg. Biochem., 99, 2197.
- (a) Fitzmaurice, R.J., Kyne, G.M., Douheret, D. and Kilburn, J.D. (2002) J. Chem. Soc. Perkin Trans., 1, 841; (b) Zhang, X.X. Bradshaw, J.S. and Izatt, R.M. (1997) Chem. Rev., 97, 3313.

- Lehn, J.M. Supramolecular Chemistry: Concepts and Perspectives, Wiley-VCH, Weinheim, 1995.
- (a) Izbicka, E., Wheelhouse, R.T., Raymond, E., Davidson, K.K., Lawrence, R.A., Sun, D., Windle, B.E., Hurley, L.H. and Von Hoff, D.D. (1999) *Cancer Res.*, 59, 639; (b) Seenisamy, J., Bashyam, S., Gokhale, V., Vankayalapati, H., Sun, D., Siddiqui-Jain, A., Streiner, N., Shin-Yan, K., White, E., Wilson, W. D. and Hurley, L.H. (2005) *J. Am. Chem. Soc.*, 127, 2944.
- Hosseni, M.W., Lehn, J.M., Duff, S.R., Gu, K. and Mertes, M.P. (1987) J. Org. Chem., 52, 1662.
- Sigman, D.S., Graham, D.R. and Aaurora, V. D. (1979) J. Biol. Chem., 254, 1269.
- 7. Downey, V.M., Que, B.R. and So, A.G. (1980) *Biochem. Biophys. Res. Commun.*, 93, 264.
- Marshall, E.L., Graham, D.R. and Reith, K.A. (1981) *Biochemistry.*, 20, 224.
- 9. Sigel, H. and Sigel, A., Eds., *Metal ions in biological systems*, Vols. 26-32, Marcel Dekker: New York, 2000.
- 10. Helleman, I. and Stock, C.C. (1983) J. Biol. Chem., 125, 771.
- 11. Yang, Z.S., Wang, Y.C. and Zhao, Z.C. (2004) Anal. Sci., 20, 1127.
- 12. Palaniandavar, M. and Ramakrishnan, S. (2005) *J. Chem. Sci.*, 117, 179.
- 13. Pradhan, R., Thomas, A.M., Mukherjee, A., Dhar, S., Nethaji, M. and Chakravarty, A.R. (2005) *Indian J. Chem.*, 44A, 18.
- (a) Prakash Naik, H.R., Bhojya Naik, H.S., Ravikumar Naik, T.R., Raja Naika, H., Gouthamchandra, K., Mahmood, R. and Khadeer

Ahamed, B.M. (2009) *Eur. J. Med. Chem.*, 44, 981; (b) Prakash Naik, H.R., Bhojya Naik, H.S., Ravikumar Naik, T.R., Raja Naik, H., Lamani, D.S. and Aravinda, T. (2008) *J. Sulfur Chem.*, 29, 583; (c) Prakash Naik, H.R., Bhojya Naik, H.S., Ravikumar Naik, T.R., Bindu, P.J., Aravinda, T. and Lamani, D.S. (2009) *Medicinal Chem.*, 5(2), 147; (d) Prakash Naik, H.R., Bhojya Naik, H.S., Ravikumar Naik, T.R., Raghavendra, M., Aravinda, T. and Lamani, D.S. (2009) *Phosphorus Sulfur Silicon and Realt. Elem.*, 184, 460.

- 15. Dhar, S., Senapati, D., Das, P.K., Chattopadhyay, P., Nethaji, M. and Chakravarty, A.R. (2003) J. Am. Chem. Soc., 125, 12118.
- Campbell, V.D., Parsons, E.J. and Pennington, W.T. (1993) *Inorg. Chem.*, 32, 1773.
- Drew, M.G.B., Esho, F.S. and Nelson, S.M. (1983) J. Chem. Soc. Dalton Trans., 1653.
- 18. Mandal, S.K. and Nag, K. (1986) J. Org. Chem., 51, 3900.
- 19. Feig, D.I., Reid, T.M. and Loeb, L.A. (1994) Cancer. Res., 54, 1890.
- 20. Routledge, M.N., Wink, D.A., Keefer, L.K. and Dipple, A. (1994) *Chem. Res. Toxicol.*, 7, 628.
- Addison, A.W., Palaniandavar, M., Driessen, W.L., Paap, F. and Reedijk, J. (1988) *Inorg. Chim. Acta.*, 142, 95.
- Pandiyan, T., Palaniandavar, M., Lakshminarayanan, M. and Manohar, H. (1992) J. Chem. Soc., Dalton Trans., 3377.
- 23. Pogozelski, W.K. and Tullius, T.D. (1998) Chem. Rev., 98, 1089.
- Maheswari, P.U., Barends, S., Zalp-Yaman, S.O., de Hoog, P., Casellas, H., Teat, S.J., Massera, C., Lutz, M., Spek, A.L., vanWezel, G.P., Gamez, P. and Reedijk, J. (2007) *Chem. Eur. J.*, 13, 5213.